

## Supplementary Information

### Bioactive Bioflavonoids from *Platonia insignis* (Bacuri) Residues as Added Value Compounds

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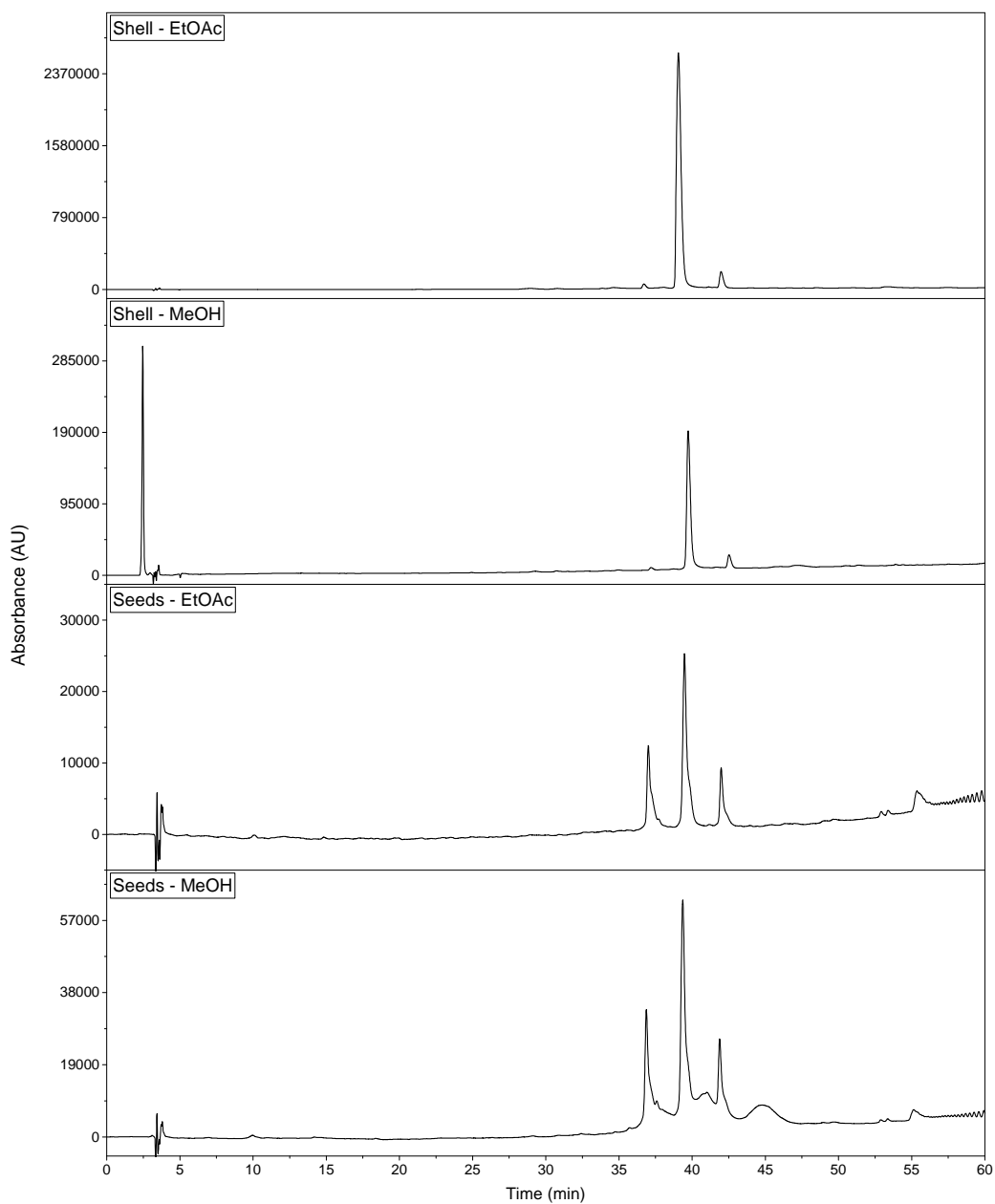
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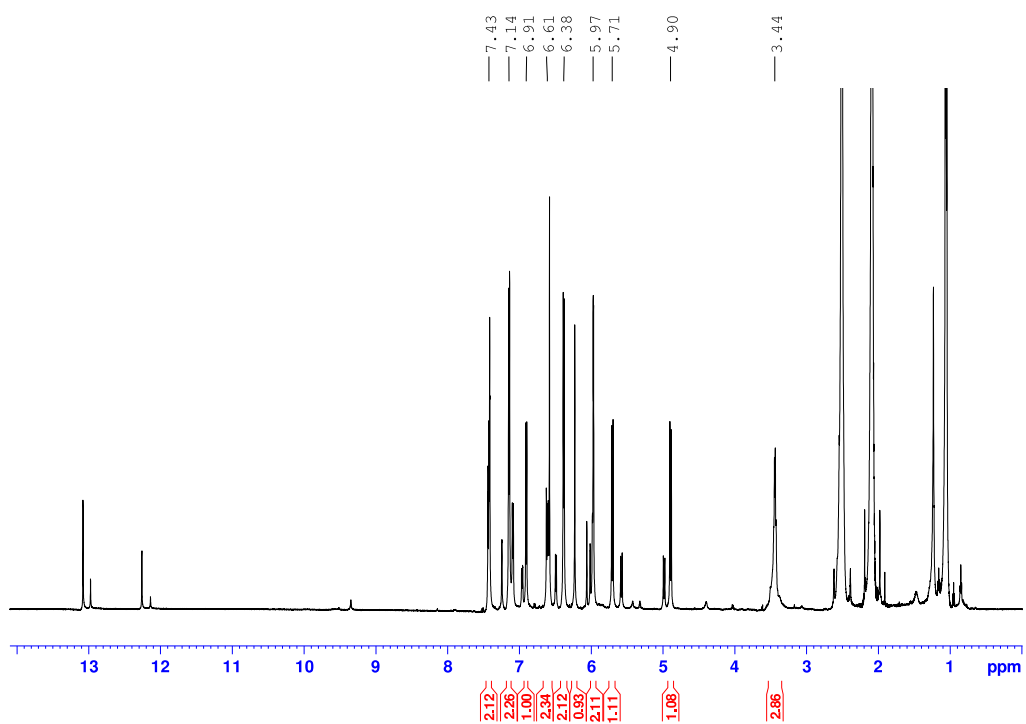
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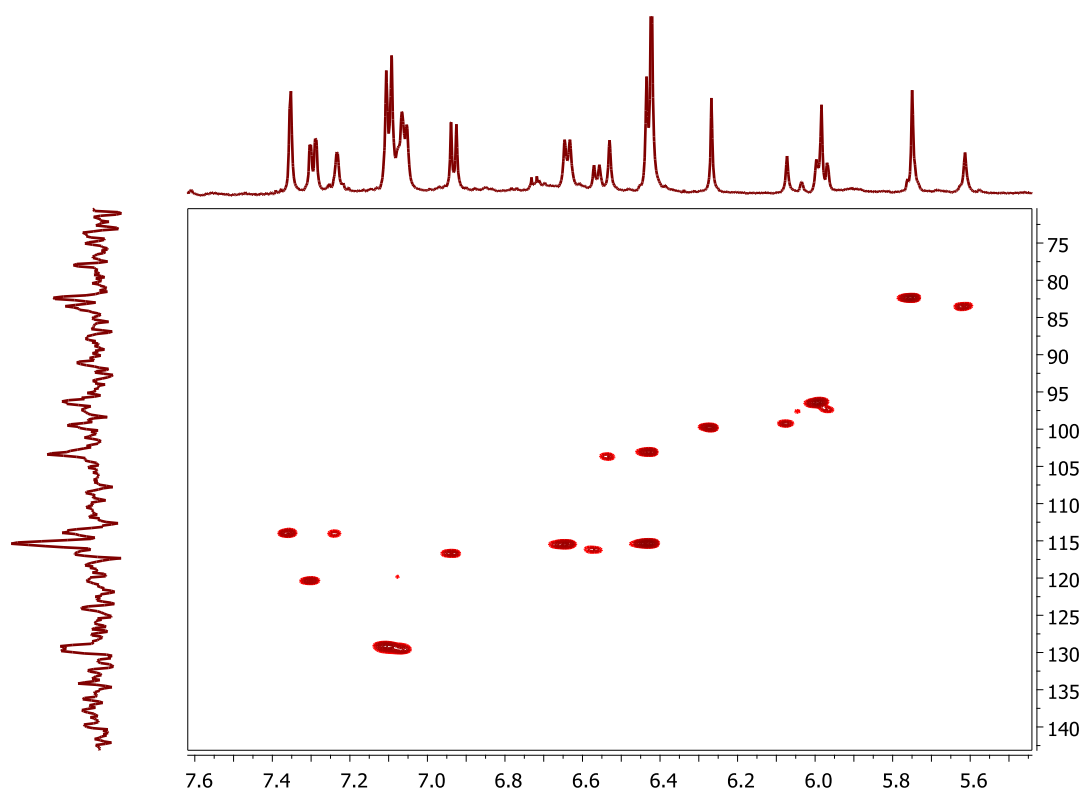
#These authors contributed equally to this work.



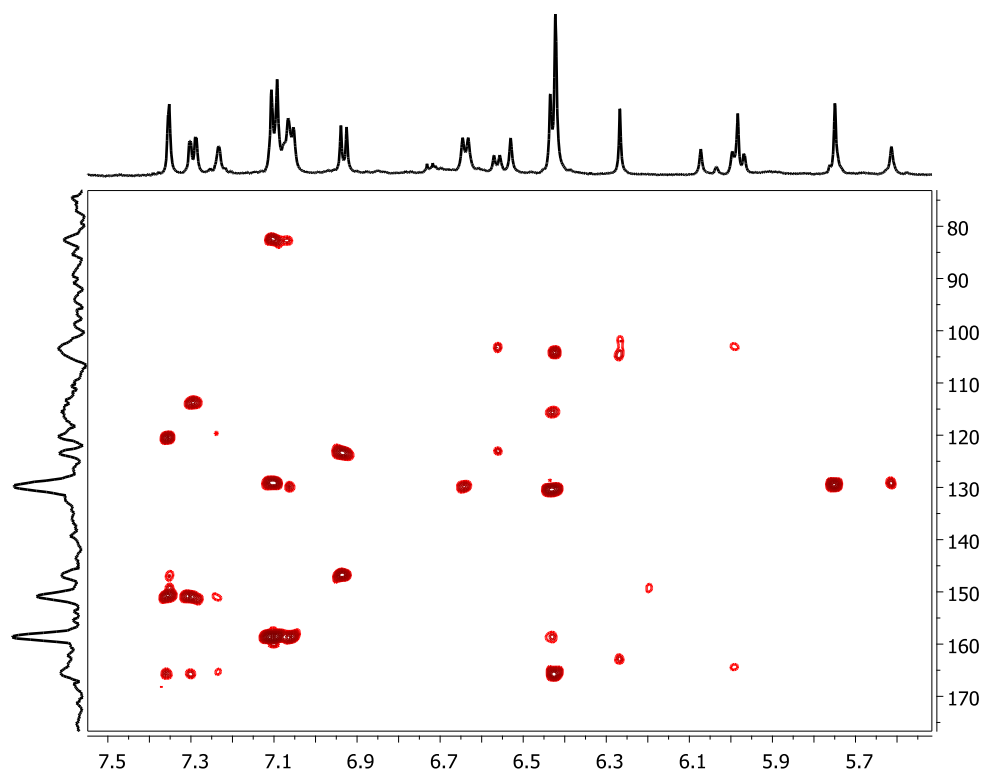
**Figure S1.** HPLC-DAD chromatograms obtained from the extracts. Chromatographic conditions: Phenomenex® C18-Luna column (250 × 4.6 mm i.d., 5 μm); 5 to 100% MeOH in 60 min; 1.0 mL min<sup>-1</sup> flow rate; 25 °C; detection at 254 nm. Samples at 10 mg mL<sup>-1</sup>; 20 μL injection volume.



**Figure S2.**  $^1\text{H}$  NMR (600 MHz,  $\text{DMSO-}d_6$ ) spectrum of compound **2**, morelloflavone.



**Figure S3.** HSQC NMR (600 MHz,  $\text{CD}_3\text{OD}$ ) spectrum of compound **2**, morelloflavone.



**Figure S4.** HMBC NMR (600 MHz, CD<sub>3</sub>OD) spectrum of compound **2**, morelloflavone.

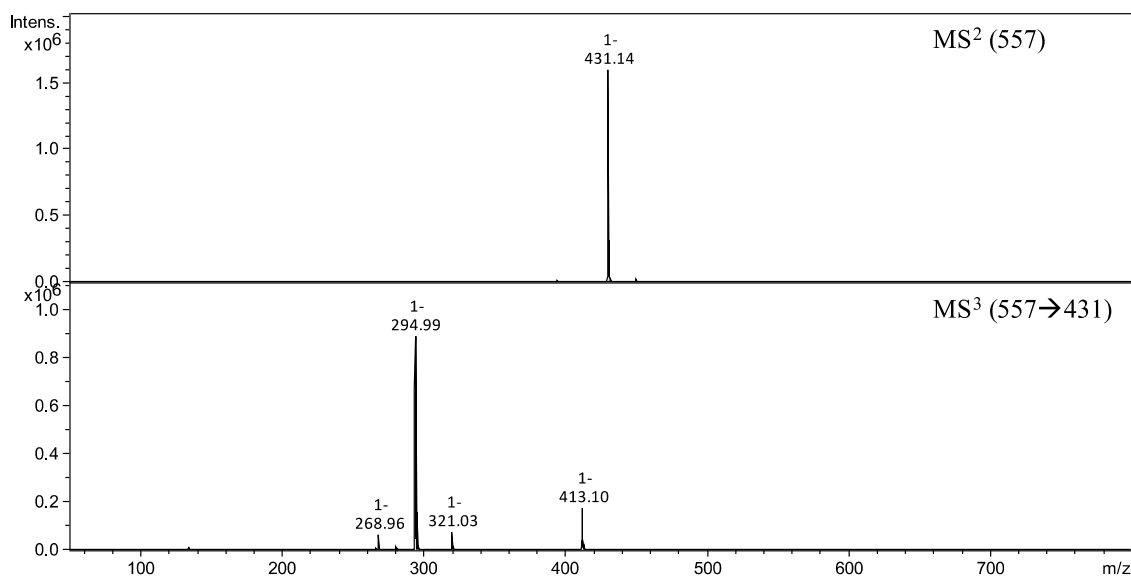
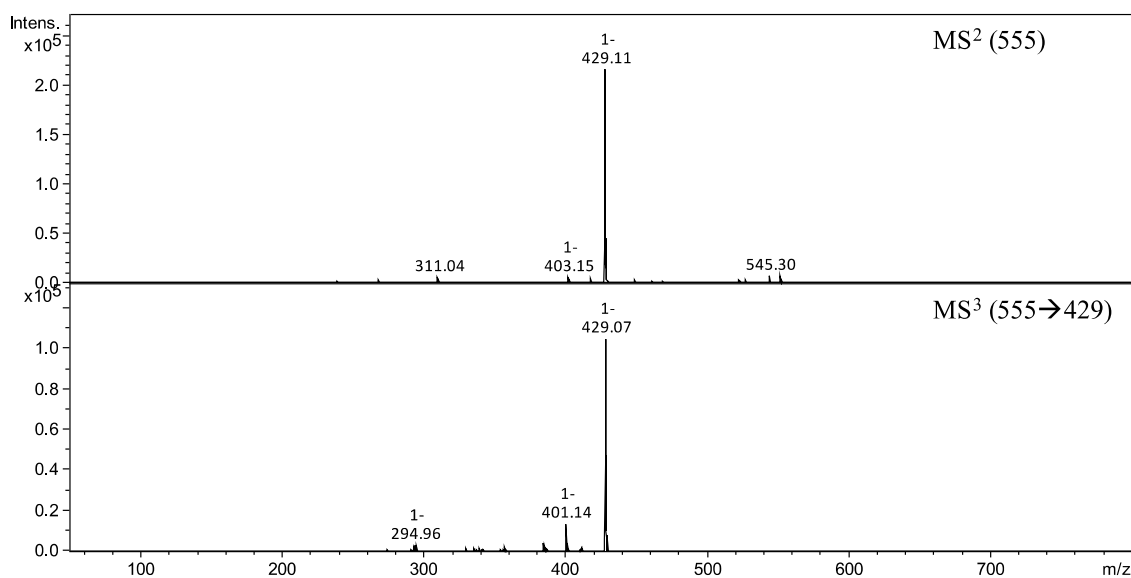
**Table S1.** Table with morelloflavone NMR chemical shifts

Position	$\delta$ <sup>1</sup> H ( <b>2a</b> ) <sup>a</sup>	$\delta$ <sup>1</sup> H ( <b>2b</b> ) <sup>a</sup>	$\delta$ <sup>13</sup> C ( <b>2a/2b</b> ) <sup>a,b</sup>	$\delta$ <sup>1</sup> H ( <b>2a</b> ) <sup>c</sup>	$\delta$ <sup>1</sup> H ( <b>2b</b> ) <sup>c</sup>
2	5.75, s <sup>d</sup>	5.61, s <sup>d</sup>	82.4/83.5	5.70, d 12.0	5.57, d 12.3
3	e	e	u/u	4.89, d 12.0	4.98, d 12.3
5-OH				12.25 s	12.13 s
6	5.99, m <sup>f</sup>	5.97, d W <sub>1/2</sub> 1.3 <sup>f</sup>	96.4/97.3	5.96, d W <sub>1/2</sub> 2.1 <sup>f</sup>	5.98, d 2.0 <sup>f</sup>
8	5.99, m <sup>f</sup>	6.04, br s <sup>f</sup>	96.4/97.6	5.97, d W <sub>1/2</sub> 2.1 <sup>f</sup>	6.01, d 2.0 <sup>f</sup>
10			129.1/129.8		
1'			129.2/129.6		
2'/6'	7.10, d 8.3	7.06, d 8.2	115.4/115.4	7.14, d W <sub>1/2</sub> 8.6	7.09, d W <sub>1/2</sub> 8.4
3'/5'	6.43, d 8.3	6.64, d 8.2	158.6/158.6	6.38, d W <sub>1/2</sub> 8.6	6.60, d W <sub>1/2</sub> 8.4
4'			165.8/165.2		
2''			103.1/103.7		
3''	6.42, s	6.53, s	183.9/u	6.58, s	6.62, s
4''					
5''-OH			99.8/99.3	13.07, s	12.97, s
6''	6.27, s	6.07, s	102.0/102.0 <sup>f</sup>	6.22, s	6.06, s
8''			104.2/u <sup>f</sup>		
10''			114.0/114.1		
2'''	7.35, d 1.7	7.23, br s	146.7/u	7.41, d W <sub>1/2</sub> 2.2	7.24, d 2.3
3'''			150.8/151.0		

**Table S1.** Table with morelloflavone NMR chemical shifts (cont.)

Position	$\delta$ $^1\text{H}$ ( <b>2a</b> ) <sup>a</sup>	$\delta$ $^1\text{H}$ ( <b>2b</b> ) <sup>a</sup>	$\delta$ $^{13}\text{C}$ ( <b>2a/2b</b> ) <sup>a,b</sup>	$\delta$ $^1\text{H}$ ( <b>2a</b> ) <sup>c</sup>	$\delta$ $^1\text{H}$ ( <b>2b</b> ) <sup>c</sup>
4 <sup>'''</sup>			116.7/116.2		
5 <sup>'''</sup>	6.93, d 8.4	6.56, d 8.3	120.4/119.8	6.90, d W <sub>1/2</sub> 8.3	6.49, d 8.5
6 <sup>'''</sup>	7.30, dd 8.4, 1.7	7.07, comp	82.4/83.5	7.43, dd W <sub>1/2</sub> 8.3, 2.2	6.96, dd 8.5, 2.3

Compounds **2a** and **2b** (intensity ratio of 1:0.35 at 25 °C, respectively) are rotamers of morelloflavone. <sup>a</sup>CD<sub>3</sub>OD; <sup>b</sup>chemical shifts of carbon nuclei were assigned with assistance of gHSQC and gHMBC experiments; <sup>c</sup>DMSO-*d*<sub>6</sub>; <sup>d</sup>signals observed as singlets in PRESAT experiment; <sup>e</sup>signals suppressed by presaturation; <sup>f</sup>assignments in the same column can be interchanged. u: undetermined chemical shifts; comp: complex multiplicity. Positions whose values of carbon chemical shifts could not be unequivocally attributed were omitted from this table.

**Figure S5.** GB-2a (**1**) MS/MS fragmentation pattern.**Figure S6.** Morelloflavone (**2**) MS/MS fragmentation pattern.

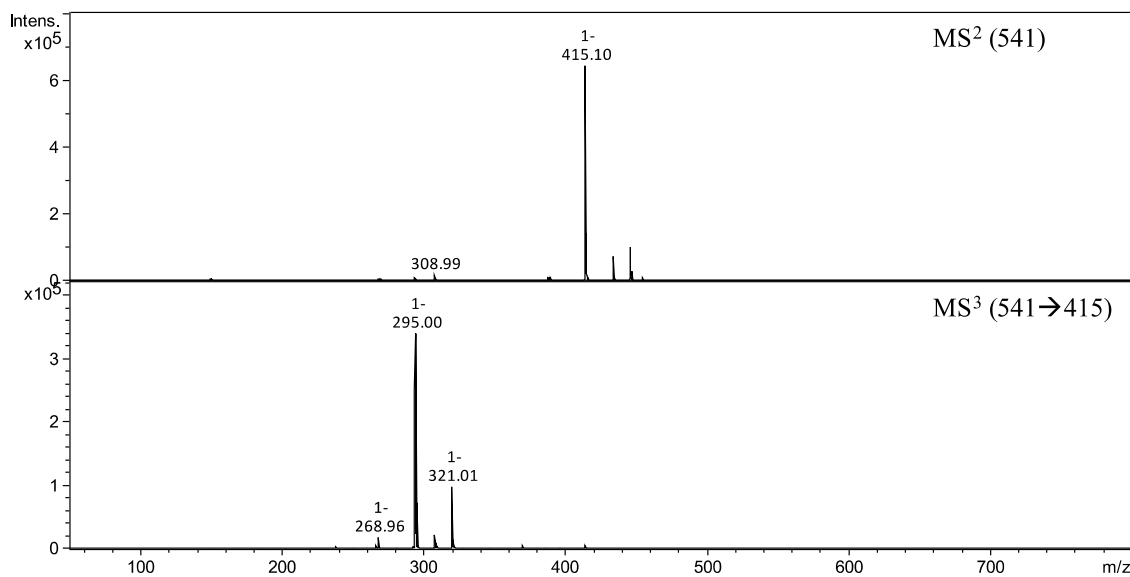


Figure S7. GB-1a (3) MS/MS fragmentation pattern.

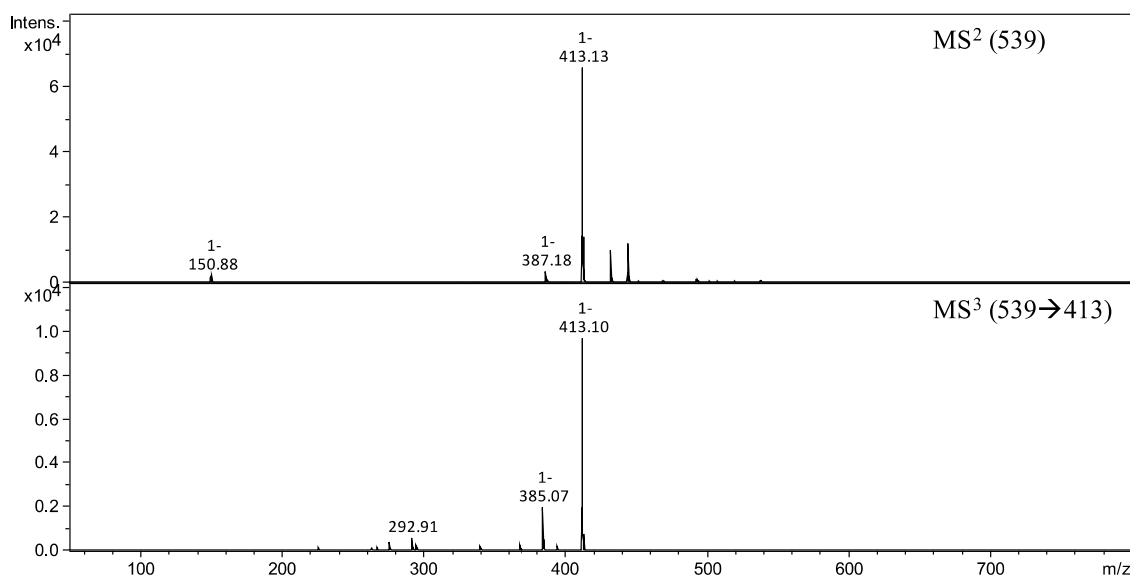
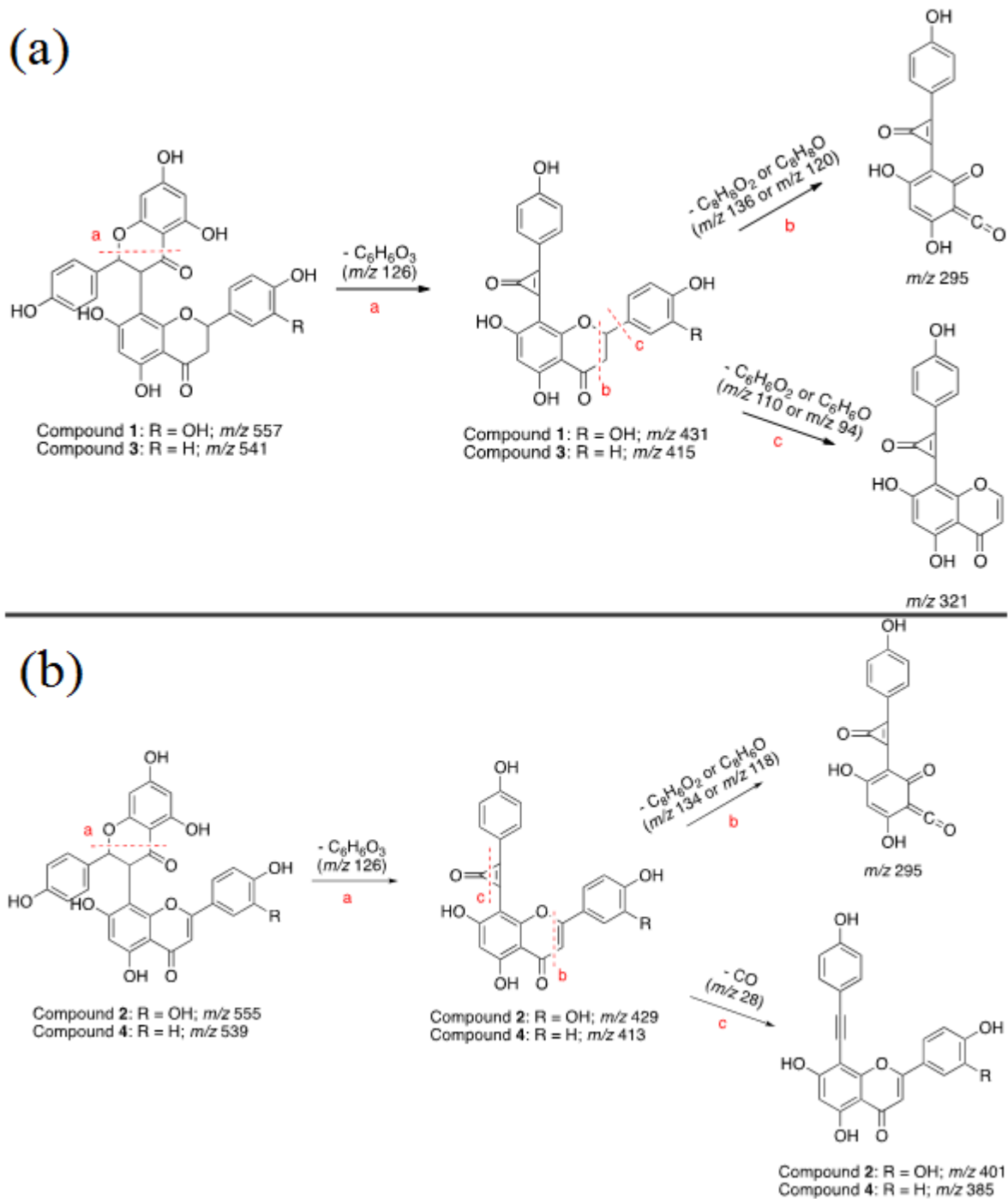
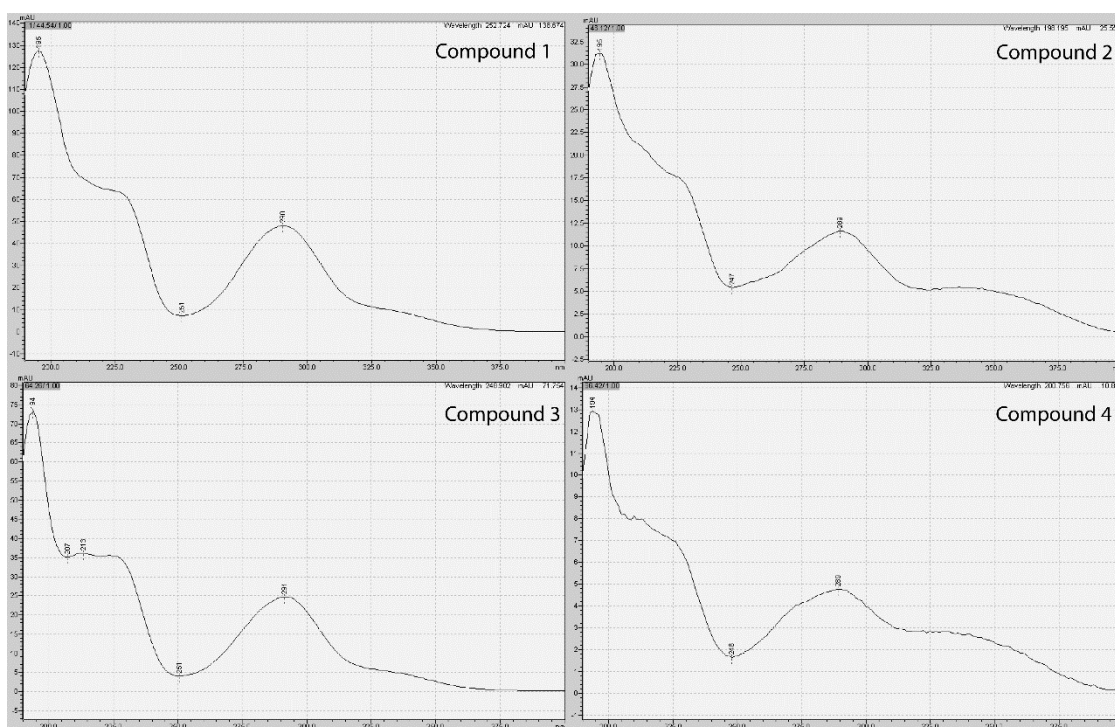


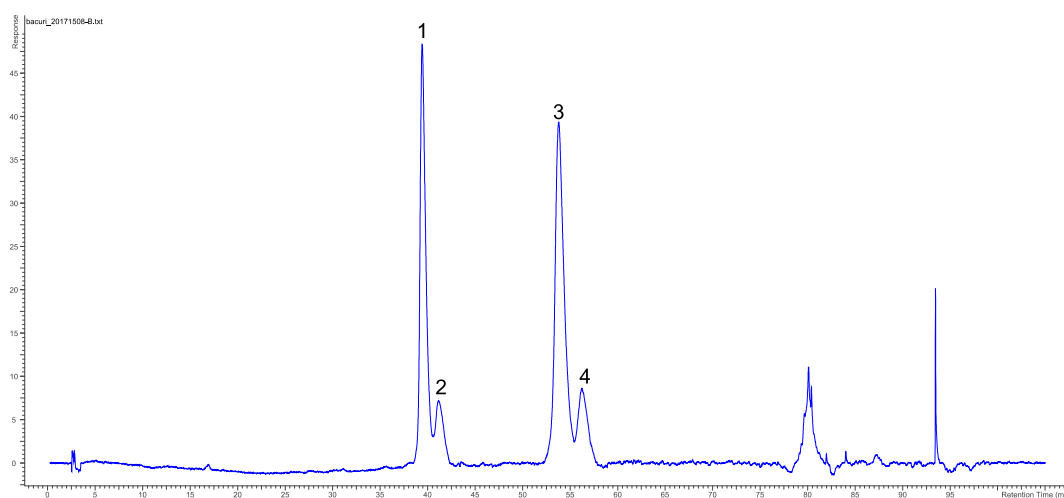
Figure S8. Volkensiflavone (4) MS/MS fragmentation pattern



**Figure S9.** MS<sup>n</sup> fragmentation pathway proposed for GB-1a (a) and morelloflavone (b) type biflavonoids. Adapted from Carrillo-Hormaza *et al.*<sup>1</sup>

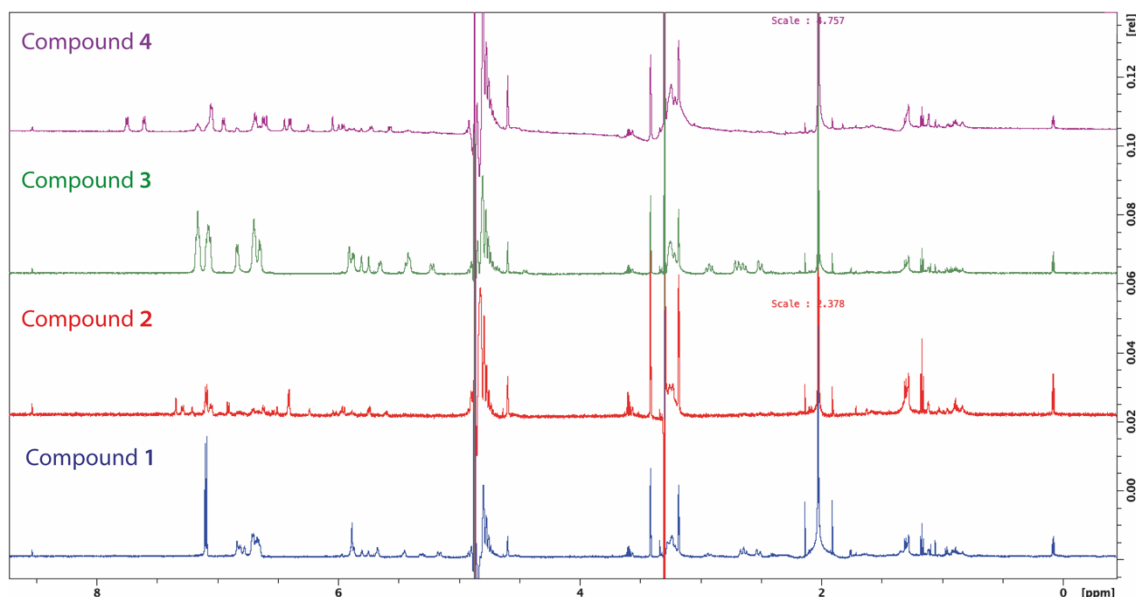


**Figure S10.** UV spectra obtained for compounds 1-4, present in *P. insignis* shell.

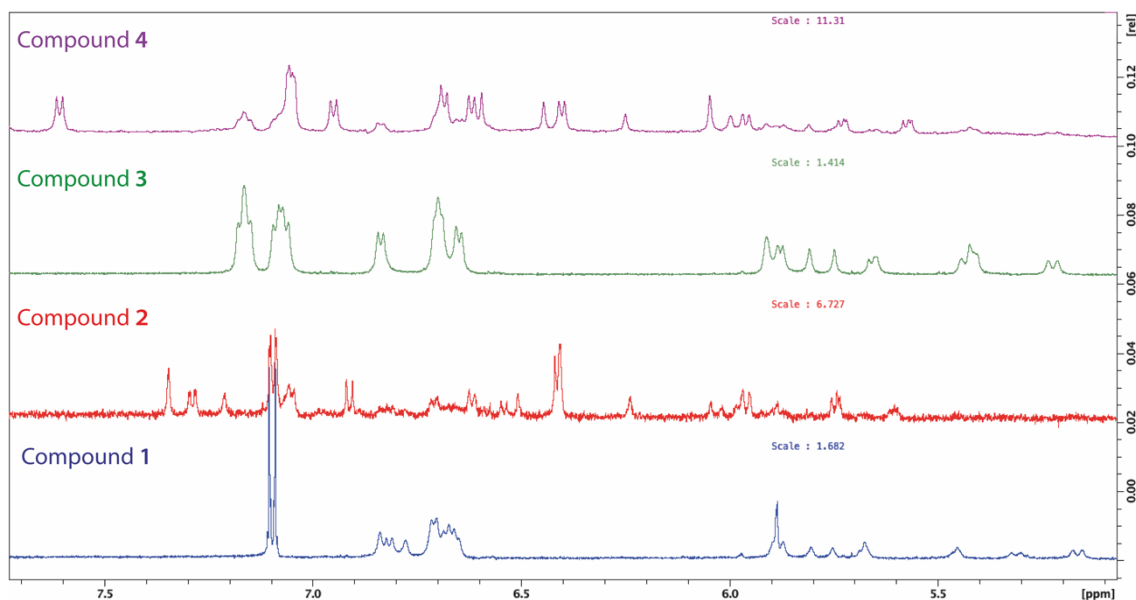


**Figure S11.** Chromatogram obtained from the LC-SPE fractionation step. Chromatographic conditions: Phenomenex® C18-Luna column (250 × 4.6 mm i.d., 5 μm); linear gradient from 5 to 30% MeOH in 25 min, followed by an isocratic mode at 30% MeOH until 75 min; 1.2 mL min<sup>-1</sup> flow rate; 25 °C; detection at 254 nm. Sample at 20 mg mL<sup>-1</sup>; 20 μL injection volume.

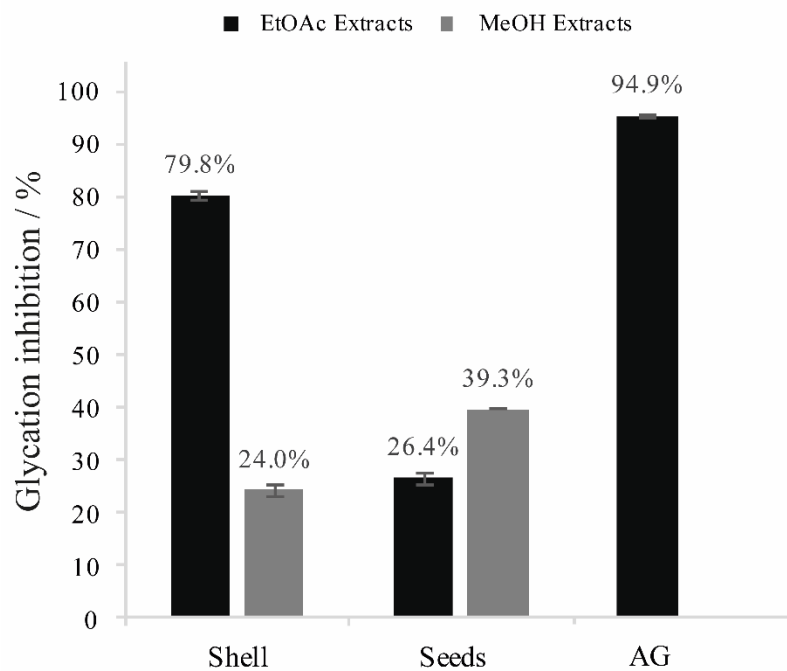




**Figure S12.** Comparison of  $^1\text{H}$  NMR with double solvent pre-saturation suppressing any residual water and methanol signals (600 MHz,  $\text{CD}_3\text{OD}-d_4$ ) spectra of compounds **1-4** obtained by LC-SPE fractionation.



**Figure S13.** Comparison of  $^1\text{H}$  NMR with double solvent pre-saturation suppressing any residual water and methanol signals (600 MHz,  $\text{CD}_3\text{OD}-d_4$ ) spectra of compounds **1-4** obtained by LC-SPE fractionation: expansion from 5.0 to 7.7 ppm.



**Figure S14.** Percentage of glycation inhibition of EtOAc and MeOH shell and seeds extracts (at  $150 \mu\text{g mL}^{-1}$ ) of *Platonia insignis*. Aminoguanidine (AG) ( $10 \text{ mmol L}^{-1}$ ) was used as positive control. Experiments were performed in triplicate.



**Figure S15.** Microscopy of polarized light for the developed crystalline system with incorporation of *P. insignis* EtOAc shell extract.

## Reference

1. Carrillo-Hormaza, L.; Ramírez, A. M.; Quintero-Ortiz, C.; Cossio, M.; Medina, S.; Ferreres, F.; Gil-Izquierdo, A.; Osorio, E.; *J. Funct. Foods* **2016**, *27*, 503.